BSES Limited



BURDEKIN NITROGEN PROJECT

BSES TECHNICAL REPORT TE11004

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PREPARED BY: JULIAN CONNELLAN¹ SUSIE BATEMAN¹ and BERNARD SCHROEDER²

¹ Research Agronomist, BSES Limited, Brandon ² Principal Scientist, BSES Limited, Bundaberg

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CONTENTS

Page Number

1. Overview	.1
2. Current Progress	.2
3. Appendix 1. Trial Protocol	.5
4. Appendix 2. PowerPoint Presentations	.14

1. Overview

In response to the introduction of the regulated method of determining N, some canegrowers in the Burdekin district (which produces approximately 25% of Queensland's sugarcane) expressed concern that sugarcane yields could be negatively affected by the permissible N application rates. Growers and Sucrogen (the milling company that operates in the Burdekin region) were also of the opinion that additional research was needed to ensure that any regulated N application would not compromise the profitability and/or supply security of the local sugarcane industry.

As a result of grower and industry concerns the Department of Environment and Resource Management (DERM) contracted BSES Limited to undertake a series of trials across the Burdekin district to determine the adequacy of the regulated N for sugarcane production on different soil types and at different on-farm locations. To date BSES has established ten trials, five of which are located in the Delta and five in the Burdekin River Irrigation Area (BRIA).

Replicated nitrogen rate strip trials have been established in plant sugarcane blocks on commercial farms in the Delta and BRIA. These sites were selected after considering a number of factors such as block size, shape, soil uniformity, irrigation systems employed and pest control measures. Consideration was also given to yield history of the blocks.

At the conclusion of this work, partial net grower returns per hectare for each N application rate will be calculated per crop and for the complete crop cycle. The optimum N application rates that maintain the profitability of sugarcane production will be identified with due consideration to the potential impact on the environment.

2. CURRENT PROGRESS

As part of the Burdekin Nitrogen Project a number of activities have been completed to date. This information has been compiled and presented to the Technical Management Group (TMG) and the Industry Reference Group (IRG) who oversee this project. TMG and IRG members are listed in Appendix 2.

At the beginning of the project protocols were developed in partnership with DERM. These protocols provide guidelines on research activities which will take place during the term of the project (refer to Appendix 1).

Eleven potential sites were initially identified through contacts made at grower group meetings, via the chairman of the IRG and through the local cane grower organisation Canegrowers, Burdekin. Once sites were identified a site inspection was undertaken with the grower to determine if the site was a prospective candidate for the nitrogen trials.

Once suitable sites were identified they were mapped using electrical conductivity resistivity measurements (Veris 3100). These maps were used to provide a general guide to changes in soil type and salinity/sodicity levels across blocks. This information was used to develop comprehensive soil sampling strategies for each site. Soil samples were collected to a depth of 1 metre, with sub-samples from 0-10 cm, 0-20 cm, 20-40 cm, 40-60 cm, 60-80 cm and 80-100cm depths being collected (Figure 3) and analysed for nutrient status separately. All soil sampling points across each site were GPS referenced (refer to Appendix 2).

GPS referenced soil sampling points

A 'soil' map generated from the electrical conductivity / resistivity measurements showing identified soil sampling points.

Results of the soil analysis from each potential site were run through the regulated method to develop recommended fertiliser application rates for each site.

During crop establishment some growers applied additional fertiliser to their plantings. These applications were recorded and fertiliser boxes calibrated to determine quantity of fertiliser applied to the potential trial site. This information was recorded on the grower information sheet (refer to Appendix 2). Crop growth /

development and factors such as pests were monitored and recorded during the establishment phase.

From the time of planting, water samples have been collected for nutrient testing by DERM. Water sources for trial sites are either bores or channel systems. In the Delta water sources are generally bores and in the BRIA generally channels however in both areas a mixture of both water sources are sometimes used.

Information gathered from soil analysis, calibration of equipment and general observations were used to develop one page information sheets (refer to Appendix 2). These information sheets were then reviewed by the TMG to determine site suitability, design and treatments/nitrogen rates for individual sites (refer to Appendix 2).

Each trial contains three randomised replicates of three or four N treatments. The treatments include:

- Treatment 1: The regulated N application rate for the particular soil type
- Treatment 2: A lower N application rate than treatment 1
- Treatment 3: An N application rate comparable to that traditionally used by the grower
- Treatment 4: A higher N rate than treatment 3.

The outcomes of this review were then presented to the IRG for their consideration and endorsement.

Of the 11 candidate sites initially selected for the trial, one site was deemed to be inadequate due to a very poor sugarcane 'strike'. This site has been excluded from the trial. However given the location, the cooperative nature of the grower and the good practices applied by the grower, it was agreed that a trial would be established on another part of this grower's property in 2012. A total of 10 sites have been endorsed by the TMG and the IRG for 2011.

It was also agreed by the TMG and the IRG that another one or two sites will be established (under the current protocols) in the Kalamia area of the Delta in 2012. The inclusion of Kalamia sites will mean all areas of the Delta are covered by the Burdekin Nitrogen Project (refer to Appendix 2 for map showing location existing and potential trial sites).

Site and treatment information was provided to the BSES biometrician to design a randomised trial setup for each of the ten sites.

At the time of this report all ten sites are in the process of being pegged-out into strips to allow for the final fertiliser application of the various nitrogen rates.

The measured application of fertiliser will take place shortly after the delivery of a DERM-funded fertiliser box that is currently being built by Gessner. Expected delivery date is late September.

Once the strip trials have been established the following will be determined and monitored over the duration of this project:

- General plant growth and pest pressure
- Plant nutrient status via leaf analyses
- Sugarcane biomass accumulation
- Irrigation water quality
- Soil moisture status (at selected sites)
- Soil nitrate values to depth
- N losses via denitrification (at selected sites)
- Sugarcane yield, commercial cane sugar (CCS) content and calculated sugar yield

Satellite imagery (to be provided by DEEDI) will be utilised to capture images of each trial site. This technology has the potential to identify differences in crop responses to the various nitrogen treatments prior to harvest.



Trial site marked with pegs identifying trial site location and fertiliser application rate

3. APPENDIX 1

9/08/2011

Burdekin Nitrogen Trials

Protocol

Partial nitrogen budgets for replicated strip trials

Components of the budget

Nitrogen inputs, outputs and status components of a budget are indicated in Fig. 1.

Direct measurements of components A, B, C and D allow a 'partial' budget to be calculated as:

Partial budget = Inputs – Outputs = (D-A) + B-C

Unless the input processes (mineralisation) and loss processes (denitrification, runoff, sediments, leaching) are measured directly, their cumulative result is reflected in the (D-A) term of the partial budget.

A net positive partial budget indicates accumulation in the soil if D>A or unmeasured net losses are occurring and are not reflected in the soil pools measured at times A and D. A net negative partial budget indicates a reduction in soil nutrient reserves which should be reflected by D<A. If D is not less than A then the soil pools measured at times A and D are not the ones providing available nutrients to the crop.





Methods for measuring components of a partial N budget

Components A and D

Total N, mineral N (ammonium-N + nitrate-N), potentially mineralisable N @ 7d, 14d of 0-20 cm, 20-40 cm, 40-60 cm, 60-80 cm, and 80-100 cm layers calculated as kg/ha using soil bulk density.

Component B

Inorganic N applied (kg/ha) as fertiliser, and nitrate-N applied in irrigation water.

Component C

Total N calculated as kg/ha using yield and %N content of above ground biomass and its moisture content.

Time of sampling components

Component A

Immediately prior to planting or immediately prior to application of fertiliser/amendment to ratoon. Any trash is removed from the soil surface before sampling.

Component C

Above-ground biomass samples will be taken at 6 months (stalk elongation) from a designated length of row for each rep, and weight of millable stalk and green leaf plus cabbage determined, with subsamples taken for total N and P analysis as detailed in 'Section 2. Protocols for establishing replicated strip trials.'

Immediately prior to harvest, another set of above-ground biomass samples will be taken as described in 'Section 2. Protocols for establishing replicated strip trials,' weighed fresh, and a subsample taken, dried at 60°C, re-weighed, ground and sub-sampled for analysis.

Component D

Immediately after harvest, collect separate composite top 0-20 cm, 20-40 cm, 40-60 cm, 60-80 cm and 80-100 cm layers calculated as kg/ha using soil bulk density from row area where biomass samples were taken. Remove any trash from soil surface before sampling.

Protocols for establishing replicated strip trials

A series of replicated demonstration strip trial sites are to be established in the Burdekin by BSES with a total of 9 sites to be established. Six sites located in the Delta and three located in the Burdekin River Irrigation Area (BRIA) and one site located at Babinda. The aim of the trials is to compare a range of nitrogen (N) application rates to a nitrogen rate determined using the method regulated under the *Reef Protection Act 2010*.

The following steps should be followed to ensure there is adequate consultation with industry and to ensure all relevant data required for this project is obtained.

1. Identify potential trial sites using a consultative process.

2. Assess each site in terms of suitability (size and shape of the block, uniformity of soil type, uniformity of standing crop for ratoon site/s, pest and disease status, uniformity of irrigation system, etc.) Plots will vary from 6 to 9 rows wide. The length of these rows will vary according to block size and shape.

3. Map the potential block using Veris 3100/EM33 to allow stratification of the block into similar units for replication and to identify soil sampling positions.

4. Identify the major soil type(s) in each potential trial block.

5. Collect composite soil samples for each potential trial site:

- Sites to be broken up into 4 sample zones according to the protocol shown in Fig 2. Adjustments to zone positions and boundaries will be done to reflect the nature of the block, the occurrence of soil types and Veris/EM maps.
- 8 10 subsamples for each depth (see bullet points below) are to be taken in each zone and combined to make one composite sample per zone.
- Where practical take a composite 0-10cm sample in each sampling zone. This sample is to be air dried (see below) and sent to Bundaberg BSES with the profile samples for forwarding on to DERM for pH, EC, exch K, Colwell-P, BSES-P, total org C, total N, W-B org C analysis.
- Take 2 sets (when in clay soils) of composite 0-20cm, 20-40cm, 40-60cm, 60-80cm & 80-100cm samples in each sampling zone.
- Once collected samples are to be air dried by leaving sample bags open.

A total of 2.0kg of soil is required for each composite sample.

- 500g for Incitec Pivot lab
- 1.5kg to be sent to Bundaberg BSES

Samples sent to Incitec Pivot are to undergo the following analysis:

- 0-20 cm *Custom test 2004-003 plus sand, silt and clay.*
- 20-40 cm *Custom test 2002-200 plus sand, silt and clay*
- 40-60 cm Custom test 2003-153 plus cations, plus sand, silt and clay.
- 60-80 cm Custom test 2003-153 plus cations, plus sand, silt and clay
- 80-100 cm Custom test 2002-200 plus sand, silt and clay

Of the 1.5kg sample sent to Bundaberg 1kg will be placed in storage and the remainder sent to DERM for the following analysis:

- Analysis by ERS, DERM, Dutton Park includes:
 - Total N, total org C
 - Mineral N
 - Potentially mineralisable N (PMN) (0-20 cm only)
 - Denitrification potential and capacity (DPC) (0-20 cm only)

6. Use the method regulated under the Act and the BSES SIX EASY STEPS to determine inputs from soil test values.

7. Determine the N treatments rates in consultation with the Industry Reference Group (IRG), Technical Management Group (TMG) and each co-operator to establish appropriate rates for each trial site.

- N treatments will comprise 4 rates:
 - A rate determined by the method regulated under the *Environmental Protection Act 1994*.
 - A rate significantly lower than the rate determined by the method regulated under the Act.
 - A rate growers can associate with i.e. a rate they typically use
 - A high rate which is significantly higher than the industry standard.

8. Rates will be decided after consideration of site/soil specific details and will be equally spaced to allow a yield response curve to be established for the block.

9. Establish each trial using the identified treatments within randomised and replicated layouts.

10. Where possible, capacitance probes, e.g. Enviroscan, or similar equipment will be installed in the block to monitor soil water. As a minimum, rainfall should be regularly recorded using a rainfall gauge at each site.

11. At every irrigation event, a sample will be taken of the irrigation water in a Bbottle supplied on request by DERM Dutton Park. Water samples will be frozen and dispatched to the DERM waters lab for nitrate, EC and pH analyses.

12. Once strips have been established choose a representative location within each strip and mark with a stake. Take an image at this marked point down the field and then an image from above looking down on the crop whilst standing on a ladder. Images to be taken at least once a month until entry is prohibited by crop size.

13. Strips to be mapped to determine the area within using Trimble GPS. For each strip map only the area to be harvested as part of the trial. For example where there are 6 rows per strip rows 1 and 6 will be guard rows. Map rows 2 to 5 to determine area. Harvested material from rows 1 and 6 from each strip will not be included in the yield determination.

14. Plant crop (before treatments have been imposed)

Where possible collect biomass/soil samples from several trial sites approximately 3 months post planting, pre-sidedressing. Two quadrats of 10 linear metres (e.g. 2 adjacent rows by 5 m each) to be hand harvested from each zone for the plant blocks. GPS all sample points. See Example 1 below.



Example 1. Biomass/soil sampling at plant stage

Biomass/soil sampling at plant stage before establishment of strips

For trial sites with 2 zones a total of 4 biomass/soil samples will be taken, for trials sites 3 zones 6 biomass/soil samples will be taken and for trial sites with 4 zones a total of 8 biomass/soil samples will be taken.

Ratoon crop (after treatments have been imposed)

Where possible collect biomass/soil samples from several trial sites approximately 3 months postharvest and sampling date referenced with respect to split-stool/sidedress N application date. In the ratoons one quadrat of 10 linear metres of sample per strip (treatment), (a total of 12 sample quadrats per trial site); however if time and resources permit an additional quadrat sampling in each strip is desirable. GPS all sample points. See Example 2 below.



Example 2a. Minimum biomass/soil sampling at ratoon stage with established strips



Example 2b. Desirable biomass/soil sampling at ration stage with established strips

Biomass/Soil sampling points for strips at ration stage

As a minimum for each site selected for biomass/soil sampling at the ration stage a minimum of 12 biomass/soil samples will be required to sample within each strip (i.e. 4 rates x 3 reps x 1 zone). If each strip and zone is then sampled the number of samples will double for trial sites with 2 zones (i.e. 4 rates x 3 reps x 2 zones) and triple for sites with 3 zones (i.e. 4 rates x 3 reps x 3 zones).

Sites with more than two zones may require some alteration to the above protocol ensure the sampling regime remains practical.

Biomass sampling. Follow method outlined below:

Collect sugarcane biomass samples *according to the following procedure:*

- From the sampling rows (not guard rows) of a plot randomly select 2 adjacent rows and measure 5 m of each. Crop density in the selected areas should be indicative of the plot and not have gaps.
- A tiller count from the plot to be undertaken (use to calculate population (tillers/m²).
- Cut all plants at ground level and weigh fresh biomass of whole plot in the field (use this to calculate biomass t/ha). This will give total fresh weight (FW) for 2 lengths of 5 m.
- Place plants in garbage bin bags, label and zip tie.
- Back at processing point undertake partitioning of material into:
 - o Green leaf
 - Cabbage (leaf sheath and immature stalk)
 - Dead leaf (not likely to be any at this 3 month stage)
 - Millable stalk (not likely to be any at this 3 month stage).
- It may be necessary to subsample the material harvested in the field depending upon the amount harvested. If subsampling is necessary then this subsample becomes the Subsample FW.
- Once partitioning of the sample (or subsample) is complete, mulch (or cut up using secateurs) the green leaf sample and the cabbage sample and place into aluminium containers (ideally get 2 samples each, one as backup). Weigh to determine the FW of samples in each container.
- Place containers in oven at 60 °C to get a dry weight, drying may take more than 2 days. Weigh daily until a constant weight is reached.

• Send samples to DERM lab for analysis of total N and P with analytical costs met from the DERM project budget.

15. Collect third-leaf samples during the leaf-sampling season (mid-November – April) from the treatment areas in each strip-trial according to existing BSES protocol. Dried samples will be sent to Bundaberg where they will be ground. The samples will then be split: one sub-sample will be processed through the BSES Leaf Analysis Service (N determinations by wet chemical methods, other nutrients according to usual procedures), the other sub-sample will be dispatched to DERM for total N and MIR analysis. Results will be distributed to IRG and co-operators.

16. Collect sugarcane biomass samples at the same time as the leaf sample according to the following procedure:

- Randomly select 2 lengths of 5 m of crop row that, are not adjacent to each other in each plot. Crop density in the selected areas should be indicative of the plot and not have gaps.
- Cut all plants at ground level and weigh biomass of whole plot in the field (use this to calculate biomass t/ha). A stalk count from the plot will also be undertaken (use to calculate stalks/m²). Collect any surface trash from one inter-row adjoining the harvested length of crop row.
- Select 20 stalks from the hand harvested material. Partition into millable stalk (MS) and green leaf and cabbage (LC) (this is usually done by cutting between the 5th and 6th dewlaps, green leaf attached to the stalk is included in LC, dead leaf is discarded). Weigh each component in the field. Use to calculate % millable stalk. Calculate Millable yield (t/ha) = Biomass (t/ha) x % MS/100. Can also be used to determine individual stalk weights, individual millable stalk weight, etc.
- The tripod method will not be suitable for the total plot biomass, particularly at final harvest. A weigh trailer will be required.
- Select a further sub-sample from these samples (e.g., about 6 stalks), partition into MS and LC and shred each component using a mobile garden mulcher. Collect a sub-sample of the freshly mulched material and weigh (aluminium food trays are used). This should be done in the field with freshly cut stalks from all plots. Transport back to the lab. Dry at 60°C and re-weigh to determine moisture content. Use moisture content to determine total dry biomass. Send about 100g each of MS and LC subsamples to the BSES lab at Indooroopilly for analysis of total N and P.

17. Regularly assess the trials for visual differences in plant growth and pest or disease symptoms.

18. Record rainfall, irrigation details (date, time on-time off, volume), fertiliser management (rate, form, placement, date), field operations. Manual rain gauges to be erected at each site and monitored for rainfall events.

19. Harvest the trials within the growers allocated harvest cycle in collaboration with harvest contractor and mill.

20. Immediately prior to burning and harvest, select 6 consecutive (or more depending upon resources) living stalks from the top and bottom of the block from each strip (replicate), separate into plant components, weigh, chop, dry at 60°C and re-weigh to determine moisture content. Mulch and send about 100g subsamples to the BSES lab at Indooroopilly for analysis of total N and P.

21. During the harvest, on the face of the crop in a middle row of each treatment select 6 consecutive (or more depending upon resources) stalks from two to three locations along the length of the strip (replicate) weigh, chop, dry at 60°C and reweigh to determine moisture content. Mulch and send about 100g subsamples to the BSES lab at Indooroopilly for analysis of total N and P.

22. During the harvest, on the face of the crop conduct stalk counts in a middle row of each treatment. Two to three 5 metre strips of stalks to be counted down each selected row.

23. Ensure the size of the replicated strips enables yield (tonnes cane/ha) and CCS data to be collected at the mill after harvest.

24. Immediately following harvest, collect separate composite 0-20 cm, 20-40 cm, 40-60 cm, 60-80 cm & 80-100 cm samples from row area where biomass samples were taken. Air dry and dispatch to ERS, DERM, Dutton Park, for the following analyses:

- i. Total N, total org C
- ii. mineral N
- iii. potentially mineralisable N (PMN)

25. A post-harvest sampling and assessment for Praty/Pachy and the free living omnivorous and predatory nematodes for selected sites where nematodes are thought to be problematic.

25. Consult with Mark Poggio (DEEDI, Ingham) to discuss economic issues relating to the trials. Calculate the partial net grower return per hectare using a standardised 'cane payment formula' to determine the partial net return per hectare to the grower:

b. Grower partial net return = ((price of sugar x (0.009 x (ccs-4)+0.6)) x cane yield) - (cane yield x estimated harvesting costs plus levies) - (fertiliser cost) (kg/ha) – (cane yield x estimated harvesting costs plus levies)

26. Provide summaries of results to the co-operating grower TMG and IRG.

27. Continue the trials for at least 3 crop cycles.

Sampling zones according to changes in soil types

	1.			
1	Sampling zone 1	Sampling zone 2	Sampling zone 3	Sampling zone 4

Comments:

- Suited when soil changes are across the field
- 4 sample areas that run parallel to row direction



Comments:

- Suited when soil changes are down the field
- 4 sample areas that run across row direction

	3.	
	Sampling zone 1	Sampling zone 2
V	Sampling zone 3	Sampling zone 4

Figure 2. - Examples of soil sampling strategies

Comments:

- Suited to when the soil is variable across and down the field
- 4 sample quadrants

4. APPENDIX 2

PowerPoint presentations

- 1. At the outset of this project a PowerPoint presentation was developed and delivered to local growers informing them of the research which was about to take place in the district. More than half a dozen local grower group meetings were attended with presentations made at these events. This PowerPoint presentation was also delivered to the Canegrowers, Burdekin board.
- 2. A PowerPoint presentation was developed and delivered to the Industry Reference Group (IRG) and the Technical Management Group (TMG) of the Burdekin Nitrogen Project on the 12th August 2011. This presentation gave a technical summary of each site and now also includes the treatments to be imposed at each site.

Copies of these PowerPoint presentations are attached.



Project aim

To assess the suitability of the Regulated Nitrogen (N) Application Rates (based on, but different to, the SIX EASY STEPS nutrient management guidelines for sustainable sugarcane production) in the Burdekin Region.

Sustainable sugarcane production: Profitable cane production in combination with environmental responsibility.







Who does what?

BSES Limited

Responsible for undertaking the 2.5 year project under contract from DERM.

- As part of the QCROPS Group (Group Manager: Dr Peter Allsopp)
- Within the Improved Cropping Systems Program (Leader: Dr Bernard Schroeder)
- Coordinated by Julian Connellan (BSES Research Agronomist)
- Technical support: John Panitz (Extension Agronomist)
- Research Assistance: Research Officer / Technician:



Who does what?

Industry Reference Group (IRG)

- David Defranciscis (Chairman): Sugarcane grower; Director, Pioneer Cane Growers
- Jim Collins (Secretary): Manager, Canegrowers, Burdekin
- · Rod Schultz: Manager, Pioneer Cane Growers Org.
- · Panikos Spyrou: Manager, Kalamia Cane Growers
- Evan Shannon: Manager, Davco Farming
- · Greg Shannon: Extension Leader, BSES Limited
- Marian Davis/Ryan Matthews: Extension Officers BSES Limited

BSES

How will we achieve our aim?

- Working in collaboration with DERM and DEEDI, BSES will:
 - Establish strip trials in plant and ration blocks in the Delta and BRIA cane growing areas
 - Trial sites are chosen after considering factors such as block size, shape, soil uniformity, irrigation system employed, pest control measures
 - · Consideration is also given to the yield history







- Once strip trials have been established monitoring of the following parameters will occur for plant/ratoon and the next 3 crops:
 - · Plant nutrient status via leaf analysis (Dec April)
 - Biomass analysis (at same time as leaf analysis)
 - Regular visual assessments of general plant growth and pest pressure
 - · Collection of yield and CCS data from strips

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Monitoring of sites (continued)

- At selected sites the following parameters will be monitored:
 - · Soil moisture status (ie.enviroscan)
 - Bore water quality (nitrates)
 - Soil nitrate status
 - N losses (denitrification)





Burdekin Nitrogen Trials













Fertiliser rates determined using SIX EASY SIEPS/method regulated under the Act for plant cane.

Image: 6/7/2011 (Planted 3/5/2011)

Zone	N rate (kg/ha)	N rate (kg/ha)	P rate (kg/ha)	K rate (kg/ha)	S rate (kg/ha)	Zn rate (kg/ha)	Cu rate (kg/ha)	Lime (t/ha)
	DYP = 150 tonnes/ha	DYP = 180 tonnes/ha	1.		1.1.2.2.4.1.1			
1	130	160	0	100	20	0	0	0
2	130	160	0	100	20	0	0	0
3	130	160	20	100	20	0	0	0
4	140	170	20	100	25	0	0	0



	Nutrient Status	Unit	Value			
Grower 2.	Sampling Depth: 0-20cm	-	Zone 1	Zone 2	Mean	
Location: BRIA	pH (1:5 Water)	-	6.4	6.5	6.5	
Latitude: 19°59'39.90"S	Organic Carbon	%	0.4	0.29	0.35	
Longitude: 147°14'59.42"E	Nitrate Nitrogen	mg/kg	8.3	5.8	7.1	
No. of rows: 55	Phosphorus (BSES)	mg/kg	88	34	61	
Row width: 1.6 m	Phosphorus Buffer Index		23	20	22	
Trial block area: 6 ha	Potassium (Amm-acet.)	meq/100g	0.2	0.11	0.16	
Variety: KQ228	Potassium (Nitric K)	meq/100g	2.1	2.2	2.2	
QDPI Soil type: 6Dbg	Sulphate Sulphur	mg/kg	5.1	2.6	3.9	

Nutrient Status	Unit	Value			
Sampling Depth: 0-20cm		Zone 1	Zone 2	Mean	
Electrical Conductivity	dS/m	0.04	0.03	0.035	
Cation Exchange Capacity	meq/100g	4.03	2.82	3.43	
Calcium (Amm-acet.)	meg/100g	3.3	2.3	2.8	
Magnesium (Amm-acet.)	meq/100g	0.5	0.39	0.45	
Sodium (Amm-acet.)	meq/100g	0.03	0.02	0.025	
Calcium/Magnesium Ratio		6.6	5.9	6.3	
Clay	%	8	5	7	
Sand (Fine)	%	67	62	64	
Sand (Coarse)	%	12	22	17	
Silt	%	14	11	12	

Soil group/order: Non-Sodic Duplex (Group 3)





Zone	N rate (kg/ha)	N rate (kg/ha)	P rate (kg/ha)	K rate (kg/ha)	S rate (kg/ha)	Zn rate (kg/ha)	Cu rate (kg/ha)	Lime (t/ha)	Mg (kg/ha)
151	DYP = 150 tonnes/ha	DYP = 180 tonnes/ha						1000	
1	150	190	0	100	15	0	0	0	Ó
2	150	180	20	100	25	0	0	Ū	0
DYP = Di	strict Vield Potentia								





Zone	N rate (kg/ha)	N rate (kg/ha)	P rate (kg/ha)	K rate (kg/ha)	S rate (kg/ha)	Zn rate (kg/ha)	Cu rate (kg/ha)	Lime (t/ha)
	DYP = 150 tonnes/ha	DYP = 180 tonnes/ha						
1	130	160	0	80	20	0	0	0
2	140	170	0	100	25	0	0	0

Grower 3	Nitrogen rates to be applie	d at plant and ratooi	n stages
	NI Dato	Plant	Ratoon
2/08/2011	Νιτάισ	kg N/ha	kg N/ha
	Very high	250	290
State State of the second s	High	210	250
	Grower/Regulated/ 6ES	170	210
Construction of the second	Low	130	170
	6ES - SIX EASY STEPS		
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	Con the Anna Land N		

Grower 4.

Location: Delta Latitude: 19°35'27.67"S Fertiliser applied at planting: Longitude: 147°21'55.68"E 100 L/ha of FlowPhos which No. of rows: contains: Zone 1: 66 N: 7.2% Zone 4: 64 P: 14% K: 7.1% Row width: 1.52 m Trial block area: 13 ha Zn: 0.5% Variety: Q200 QDPI Soil type: BUfc Soil group/order: Dermosol



regulated under the Act for plant cane

Nutrient Status	Unit	2.2-1		Value	1	
Sampling Depth: 0-20cm	1	Zone 1	Zone 2	Zone 3	Zone 4	Mean
pH (1:5 Water)		7.0	7.0	7.0	7.1	7.0
Organic Carbon	%	0.88	1.1	1.0	0.97	0.99
Nitrate Nitrogen	mg/kg	4.0	1.6	4.0	3.1	3.2
Phosphorus (BSES)	mg/kg	130	160	160	170	155
Phosphorus Buffer Index		45	42	50	45	46
Potassium (Ammacet.)	meg/100g	0.15	0.2	0.18	0.18	0.18
Potassium (Nitric K)	meq/100g	3.6	3.8	3.6	3.3	3.6
Sulphate Sulphur	mg/kg	4.1	3.5	4.1	3.1	3.7

Fertiliser rates determined using SIX EASY STEPS/method



Nitrogen Rates (4 treatments, 72 rows) Plant: 120kg/ha, 160kg/ha, 220kg/ha 280kg/ha Ratoon: 160kg/ha, 200kg/ha, 260kg/ha 300kg/ha

Fertiliser applied on the 2/8/2011: N: 52 kg/ha S: 62 kg/ha

Nutrient Status	Unit	1		Value	(100
ampling Depth: 0-20cm		Zone 1	Zone 2	Zone 3	Zone 4	Mean
Jectrical Conductivity	dS/m	0.04	0.04	0.04	0.04	0.04
ation Exchange Capacity	meq/100g	11.2	14.1	13.7	13.0	13
acium(Ammacet.)	meg/100g	8.0	10.0	10.0	9.5	9.4
agnesium(Ammacet.)	meq/100g	2.8	37	3.3	3.2	33
odium(Ammacet.)	meg/100g	0.21	0.21	0.2	0.17	0.2
alcium/Magnesium Ratio		2.9	27	3.0	3.0	2.9
Xlay	%	16	21	20	19	19
Sand (Fine)	%	59	55	52	53	55
Sand (Coarse)	%	6	3	8	9	6
Silt	%	19	21	20	20	20

Traditional grower Nitrogen application rate for this site: 230 kg/ha

Zone	N rate (kg/ha)	N rate (kg/ha)	P rate (kg/ha)	K rate (kg/ha)	S rate (kg/ha)	Zn rate (kg/ha)	Cu rate (kg/ha)	Lime (t/ha)
	DYP = 150 tonnes/ha	DYP = 180 tonnes/ha						
1	130	160	0	100	20	0	0	0
2	130	160	0	100	20	0	0	0
3	130	160	0	100	20	0	0	0
4	130	160	0	100	20	0	0	0

DYP = District Yield Potential



Grower 5. Nutrient Status Unit Value Nutrient Status Unit Value Sampling Depth: 0-20cm Zone 1 Zone 2 Zone 3 Zone 4 Mean Location: BRIA Sampling Depth: 0-20cm Zone 1 Zone 2 Zone 3 Zone 4 Mean 0.03 0.04 0.03 0.04 pH (1:5 Water) 7.2 7.1 7.1 7.2 7.2 Electrical Conductivity dS/m 0.04 Latitude: 19°34'44.97"S Organic Carbon Cation Exchange Capacity neq/100g 9.0 11.2 8.51 12.2 10.2 % 0.43 0.41 0.48 0.55 0.47 Longitude: 147°9'17.25"E Calcium (Amm-acet.) neq/100g 6.0 7.5 5.5 8.0 6.8 Nitrate Nitrogen 2.1 2.4 mg/kg <1.0 1.6 No. of rows: 93 Magnesium (Amm-acet.) meq/100g 2.7 3.4 2.7 3.8 3.2 Phosphorus (BSES) mg/kg 87 65 73 110 84 Sodium (Amm-acet.) neq/100g 0.1 0.14 0.15 0.19 0.15 Row width: 1.6 m Phosphorus Buffer Index 44 49 49 57 50 Calcium/Magnesium Ratio 2.2 2.0 2.1 2.2 2.1 Potassium (Amm-acet.) 0.16 0.19 Trial block area: 9 ha meq/100g 0.2 0.14 0.17 Clay 20 25 21 28 23 1.9 1.9 % Potassium (Nitric K) meq/100g 2.0 1.9 1.9 Variety: Q208 Sand (Fine) % 44 46 40 36 42 Sulphate Sulphur mg/kg 13 1.2 1.4 1.5 1.4 QDPI Soil type: 6Drc % Sand (Coarse) 26 14 16 12 17 % Silt Soil group/order: Sodic Duplex 18 18 16 21 18 Zone Removed from trials due to poor strike Zone 1 Zone 2 Zone 3 Image 6/7/2011 Fertiliser applied at planting: Planted: 12-16 May 2011 N: 30 kg/ha Traditional grower nitrogen application P: 9 kg/ha K: 50 kg/ha rate for this site: 210 kg/ha S: 26 kg/ha N rate (kg/ha) Prate Krate Srate Zn rate Cu rate Lime Mg rate Zone N rate (kg/ha) DYP = 150 tonnes/ha DYP = 180 tonnes/ha (kg/ha) (kg/ha) (kg/ha) (kg/ha) (kg/ha) (t/ha) (kg/ha) 1 140 170 0 100 25 0 0 0 0 2 140 170 0 100 25 10 0 0 0 3 140 170 100 25 0 0 0 0 0 4 140 170 0 100 25 0 0 0 0 DYP = District Yield Potential





Zone	N rate (kg/ha)	N rate (kg/ha)	P rate	K rate	S rate	Zn rate Cu rate		Lime	Mg rate	
12.2	DYP = 150 tonnes/ha	DYP = 180 tonnes/ha	(kg/ha)	(kg/ha)	(kg/ha)	(kg/ha)	(kg/ha)	(t/ha)	(kg/ha)	
1	140	170	20	100	15	10	0	0	0	
2	140	170	30	50	15	10	0	0	0	
DYP = I	District Yield Potential						1			



	Nutrient Status	Unit	10.77	Value		Later Banks			
Carrier 7	Sampling Depth: 0-20cm	1.00	Zone 1	Zone 2	Mean	MATSON I WE AND A MARKED			
Grower 7.	pH (1:5 Water)	1	7.8	8.3	8.1	5-2-5-5-711	a a		
Location: Delta	Organic Carbon	%	0.51	0.64	0.58	and the second second	La Maria		
Latitude: 19°42'8.46"S	Nitrate Nitrogen	mg/kg	2.3	2.5	2.4		A Cart	Stor y	
Longitude: 147°25'3.66"E	Phosphorus (BSES)	mg/kg	25	17	21	1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	からう	1.4.4	1
No. of rows: 80	Phosphorus Buffer Index		60	100	80		A. T. D.	a sale	
Row width: 1.52 m	Potassium (Amm-acet.)	meq/100g	0.19	0.18	0.19	Imaga 6/7/2011	THE R. P.	1 3 3 K	5
Trial block area: 11 ha	Potassium (Nitric K)	meg/100g	2.6	2.6	2.6	image onizon			
Variety: Q208	Sulphate Sulphur	mg/kg	11.0	2.7	6.9		*	/	
Ra	toon: 170kg/ha, 210kg/ha, 250k applied at planting:	ria, 250kg/na ig/ha, 290kg/	a /ha		*5	* 5 × 7 × 15	15		
Rat Fertiliser N: 31 kg/h P: 9 kg/ha K: 55 kg/h S: 28 kg/h	toon: 170kg/ha, 210kg/ha, 250k applied at planting: ha a zone 1	ig/ha, 290kg/ha	a /ha	20	*5 me 2	Nutrient Status	Unit		Value
Rat Fertiliser N: 31 kg/h P: 9 kg/ha K: 55 kg/h S: 28 kg/h	toon: 170kg/ha, 210kg/ha, 250k applied at planting: ha a zone 1	ig/ha, 290kg/la	a /ha	20	*5 me 2	Nutrient Status Sampling Depth: 0-20cm	Unit	Zone 1	Value Zone 2
Rat Fertiliser N: 31 kg/h P: 9 kg/ha K: 55 kg/h S: 28 kg/h	toon: 170kg/ha, 210kg/ha, 250k applied at planting: ha a zone 1	rg/ha, 290kg/ha g/ha, 290kg/	a /ha	20	*5 me 2	Nutrient Status Sampling Depth: D-20cm Electrical Conductivity	Unit dS/m	Zone 1 0.07	Value Zone 2 0.06
Rat Fertiliser N: 31 kg/h P: 9 kg/ha K: 55 kg/h S: 28 kg/h	toon: 170kg/ha, 210kg/ha, 250k applied at planting: ha zone ba zone toon	rg/ha, 290kg.	a /ha	20	me 2	Nutrient Status Sampling Depth: D-20cm Electrical Conductivity Cation Exchange Capacity	Unit dS/m meq/100g	Zone 1 0.07 13.4	Value Zone 2 0.06 15.4
Rat Fertiliser N: 31 kg/h P: 9 kg/ha K: 55 kg/h S: 28 kg/h	toon: 170kg/ha, 210kg/ha, 250k applied at planting: ha a a b b cov denetion	ra, 290kg/ha, 290kg/	a /ha	20	*5 ne 2	Nutrient Status Sampling Depth: D-20cm Electrical Conductivity Cation Exchange Capacity Calcium (Amm-acet.)	Unit dS/m meq/100g meq/100g	Zone 1 0.07 13.4 8.5	Value Zone 2 0.06 16.4 10.0
Rat Fertiliser N: 31 kg/h P: 9 kg/ha K: 55 kg/h S: 28 kg/h	toon: 170kg/ha, 210kg/ha, 250k applied at planting: ha a a a cone toor dractar	rg/ha, 290kg.	a /ha	20	*5 ne 2	Nutrient Status Sampling Depth: 0-20cm Electrical Conductivity Cation Exchange Capacity Calcium (Amm-acet.) Magnesium (Amm-acet.)	Unit dS/m meq/100g meq/100g meq/100g	Zone 1 0.07 13.4 8.5 4.1	Value Zone 2 0.06 16.4 10.0 5.3
Rat Fertiliser N: 31 kg/h P: 9 kg/ha K: 55 kg/h S: 28 kg/h	toon: 170kg/ha, 210kg/ha, 250k applied at planting: ha ha ba cone to the second	ra, 290kg/ha ;g/ha, 290kg.	a /ha	20	*5 ne 2	Nutrient Status Sampling Depth: 0-20cm Electrical Conductivity Cation Exchange Capacity Calcium (Amm-acet.) Magnesium (Amm-acet.) Sodium (Amm-acet.)	Unit dS/m meq/100g meq/100g meq/100g meq/100g	Zone 1 0.07 13.4 8.5 4.1 0.65	Value Zone 2 0.06 16.4 10.0 5.3 0.91
Rat Fertiliser N: 31 kg/h S: 9 kg/ha S: 28 kg/h	toon: 170kg/ha, 210kg/ha, 250k applied at planting: ha ha Row dmmm Row dmmm	ra, 290kg/ha, 290kg.	a /ha	20	*5 me 2	Nutrient Status Sampling Depth: 0-20cm Electrical Conductivity Cation Exchange Capacity Calcium (Amm-acet.) Magnesium (Amm-acet.) Sodium (Amm-acet.) Calcium/Magnesium Ratio	Unit dS/m meq/100g meq/100g meq/100g	Zone 1 0.07 13.4 8.5 4.1 0.65 2.1	Value Zone 2 0.06 16.4 10.0 5.3 0.91 1.9
Rat Fertiliser N: 31 kg/h S: 9 kg/hs S: 28 kg/h	toon: 170kg/ha, 210kg/ha, 250k applied at planting: ha ha Row dmmm Row dmmm	ra, 290kg/ha ig/ha, 290kg.	a /ha	20	ne 2	Nutrient Status Sampling Depth: 0-20cm Electrical Conductivity Cation Exchange Capacity Calcium (Amm-acet.) Magnesium (Amm-acet.) Sodium (Amm-acet.) Calcium/Magnesium Ratio Clay	Unit dS/m meq/100g meq/100g meq/100g meq/100g	Zone 1 0.07 13.4 8.5 4.1 0.65 2.1 24	Value Zone 2 0.06 16.4 10.0 5.3 0.91 1.9 28
Rat Fertiliser N: 31 kg/h S: 9 kg/h S: 28 kg/h	toon: 170kg/ha, 210kg/ha, 250k applied at planting: ha ha Row domain Row domain	rg/ha, 290kg/ha gg/ha, 290kg/	a /ha	20	me 2	Nutrient Status Sampling Depth: 0-20cm Electrical Conductivity Cation Exchange Capacity Calcium (Amm-acet.) Magnesium (Amm-acet.) Sodium (Amm-acet.) Calcium/Magnesium Ratio Clay Sand (Fine)	Unit dS/m meq/100g meq/100g meq/100g meq/100g	Zone 1 0.07 13.4 8.5 4.1 0.65 2.1 24 50	Value Zone 2 0.06 16.4 10.0 5.3 0.91 1.9 28 47
Rat Fertiliser N: 31 kg/h S: 9 kg/h S: 28 kg/h	toon: 170kg/ha, 210kg/ha, 250k applied at planting: ha a concernent row one row one ro	al grower N	itrogen	applicat	ion	Nutrient Status Sampfing Depth: 0-20cm Electrical Conductivity Calcium (Amm-acet.) Magnesium (Amm-acet.) Sodium (Amm-acet.) Calcium/Magnesium Ratio Clay Sand (Fine) Sand (Coarse)	Unit dS/m meq/100g meq/100g meq/100g % %	Zone 1 0.07 13.4 8.5 4.1 0.65 2.1 24 50 9	Value Zone 2 0.06 16.4 10.0 5.3 0.91 1.9 28 47 5

Mean 0.07 14.9 9.3 4.7

0.78 2.0 26 48

7 19

Zone	N rate (kg/ha)	N rate (kg/ha)	P rate	K rate	S rate	Zn rate	In rate Cu rate		Mg rate	
	DYP = 150 tonnes/ha	DYP = 180 tonnes/ha	(kg/ha)	(kg/ha)	(kg/ha)	(kg/ha)	(kg/ha)	(t/ha)	(kg/ha)	
1	140	170	20	100	10	0	0	0	0	
2	140	170	30	100	25	10	0	0	0	
DYP = [District Yield Potential									





Zone	N rate (kg/ha)	N rate (kg/ha)	P rate	K rate	S rate	Zn rate	Cu rate	Lime	Mg rate
1	DYP = 150 tonnes/ha	DYP = 180 tonnes/ha	(kg/ha)	(kg/ha)	(kg/ha)	(kg/ha)	(kg/ha)	(t/ha)	(kg/ha)
1	140	170	30	80	10	0	0	0	0
2	140	170	40	80	0	0	0	0	0
DYP = I	District Yield Potential		1	1.00		1		12 . A. 1	1

Fertiliser rates determined using SIX EASY STEPS/method regulated under the Act for plant cane.





DYP = District Yield Potential





Fertiliser rates for each zone are	determined using SIX EAS	SY STEPS/method rea	gulated under the Act for a	plant cane.

Zone	N rate (kg/ha)	N rate (kg/ha)	P rate	K rate	S rate	Zn rate	Cu rate	Lime	Mg rate
	DYP = 150 tonnes/ha	DYP = 180 tonnes/ha	(kg/ha)	(kg/ha)	(kg/ha)	(kg/ha)	(kg/ha)	(t/ha)	(kg/ha)
1	140	170	30	80	25	10	0	0	0
2	130	160	20	80	10	10	0	0	0
3	140	170	0	50	25	10	0	0	0



Grower 11 Location: BRIA Latitude: 19°41'20.90"S Longitude: 147°13'19.21 No. of rows: 144	"E		Fertiliser applied at planting: N: 38 kg/ha P: 43 kg/ha K: 36 kg/ha S: 19 kg/ha	Fertiliser applied on the 13 th July: N: 33 kg/ha S: 64 kg/ha	Traditional grower Nitrogen application rate for this site: 180kg/ha
Row width: 1.5 m (Contr Trial Block Area: 21 hec Variety: KQ228 QDPI soil type: 6Drc Soil group/order: Sodic I	olled traff tares Duplex	īc)		004	Image 1/8/2011 Planted 5th May 2011
Nutrient Status	Unit	Value			
Sampling Depth: 0-20cm					The second second second second second
pH (1:5 Water)	-	7.3	374 1	1 100	A REAL PROPERTY AND A REAL
Organic Carbon	96	0.9	Pro la construcción de la constr		
Nitrate Nitrogen	mg/kg	11		1	A CONTRACT OF THE OWNER OF THE
Phosphorus (BSES)	mg/kg	73			THE PARTY OF THE STREET OF THE
Phosphorus Buffer Index		103.7	AN A REAL PROPERTY OF A REAL PRO	1	
Potassium (Amm-acet.)	meq/100g	0.47	M IN SHEEP IN STREET		
Potassium (Nitric K)	meq/100g	2.49		1	
Sulphate Sulphur	mg/kg	5.1	A life the second	1	
Nutrient Status	Unit	Value			Nitrogen Rate (3 treatments x 16
Sampling Denth: 0-20cm	- Crim	valu			Tows/treament =144 Tows
Electrical Conductivity (15 k20)	dSim	0.08			Plant: 130kg/ha, 170kg/ha, 210kg/ha
ECEC	men/100r	20.00	-		Ratoon: 170kg/ha, 210kg/ha, 250kg/ha
Calcium (Amm.acet)	men/100g	1 12 1			Contract A variante of the second second second
Magnesium (Amm-acet)	meg/100c	74		the second se	
Sodium (Amm.acet.)	meg/100g	0.9		The state of the s	
Sodium % of Cations (ESP)		43	DO2	in Phy L18	
opplain is of equally (For)	14	14.0	- Constant - Constant		

N rate (kg/ha)	N rate (kg/ha)	P rate	K rate	S rate	Zn rate	Cu rate	Lime	Mg rate
DYP = 150 tonnes/ha	DYP = 180 tonnes/ha	(kg/ha)	(kg/ha)	(kg/ha)	(kg/ha)	(kg/ha)	(t/ha)	(kg/ha)
130	160	0	0	20	0	0	0	0
DYP = District Yield Potential			1				1	A



